

Evolution of the Z-scheme of photosynthesis: a perspective

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Received: 7 November 2016 / Accepted: 29 December 2016
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Abstract The concept of the Z-scheme of oxygenic photosynthesis is in all the textbooks. However, its evolution is not. We focus here mainly on some of the history of its biophysical aspects. We have arbitrarily divided here the 1941–2016 period into three sub-periods: (a) Origin of the concept of two light reactions: first hinted at, in 1941, by James Franck and Karl Herzfeld; described and explained, in 1945, by Eugene Rabinowitch; and a clear hypothesis, given in 1956 by Rabinowitch, of the then available cytochrome experiments: one light oxidizing it and another

reducing it; (b) Experimental discovery of the two light reactions and two pigment systems and the Z-scheme of photosynthesis: Robert Emerson's discovery, in 1957, of enhancement in photosynthesis when two light beams (one in the far-red region, and the other of shorter wavelengths) are given together than when given separately; and the 1960 scheme of Robin Hill & Fay Bendall; and (c) Evolution of the many versions of the Z-Scheme: Louis Duysens and Jan Ames's 1961 experiments on oxidation and reduction of cytochrome *f* by two different wavelengths of light, followed by the work of many others for more than 50 years.

Submitted for publication in honor of Nathan Nelson, a world leader in the field of photosynthesis, and of T. Nejat Veziroglou, a world leader in the field of hydrogen evolution (see Tsygankov et al. 2016; also see pdfs at <http://www.life.illinois.edu/govindjee/honorsfrom.html>).

John Raven sent the following comment on this paper: "The history of the concept of the 'Z scheme' analysed in this manuscript by Govindjee and colleagues is timely and accurate. The sequence of publications cited, and the discussion of these publications, show how theoretical and experimental work led to our present concept of linear electron transport in oxygenic photosynthesis".

Electronic supplementary material The online version of this article (doi:10.1007/s11120-016-0333-z) contains supplementary material, which is available to authorized users.

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Keywords Louis N. M. Duysens · Robert Emerson · James Franck · Robin Hill · Bessel Kok · Eugene Rabinowitch · Horst T. Witt · The Z-scheme of photosynthesis

The main purpose of this educational paper is to provide an overview of the origin of the concept of the two light reactions and two photosystems that has evolved during the last 75 years. Obviously, the evolution of this concept into the modern Z-scheme (as we know it today) did not take

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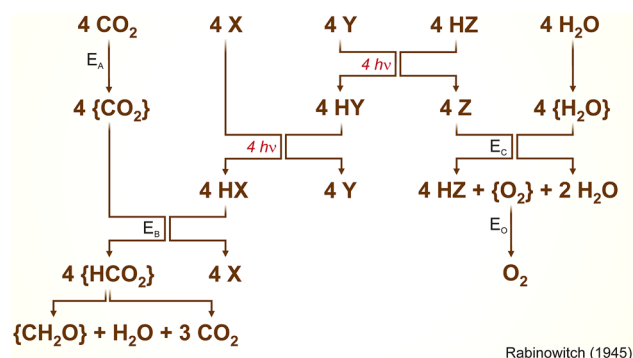
much time on a historical scale, but it required tremendous intellectual input of many research groups from all over the World. We present below our personal perspective for three time periods: 1941–1956; 1957–1960; and 1961—the present. No attempt is made here to have an all-encompassing review, but it includes some representative citations. For an earlier brief account, see Govindjee et al. (2012), and for a detailed account, see Govindjee and Björn (2015).

Origin of the concept of two light reactions (1941–1956)

Eugene I. Rabinowitch (1945) (born, b.1901–died, d.1973; see Bannister 1972) was the first to discuss the statement of 1941 by James Franck (1882–1964) and Karl F. Herzfeld (1892–1978) of two light reactions to explain the fact that a minimum of 8–12 quanta are required to evolve one oxygen molecule, or fix one CO_2 molecule (see Appendix for views on James Franck's concepts on photosynthesis—that were often in error, and at odds with the existing literature). Figure 1 shows Rabinowitch's (1945) scheme, which was considered, among many other schemes, to accommodate available results and existing ideas. Robert Emerson (1903–1959) and Charleton Lewis (see Emerson and Lewis 1943, and Rabinowitch 1961) had clearly established that the minimum quantum requirement for oxygen evolution was not 4 but 8–12. For further information on the minimum quantum requirement (or its inverse, the maximum quantum yield for oxygen evolution), and the controversy surrounding it, see Govindjee et al. (1968); Nickelsen and Govindjee (2011); and Hill and Govindjee (2014).

In the Scheme 7.V (Rabinowitch 1945, p. 162), oxidation of an unknown intermediate HZ to Z would be equivalent to today's oxidation of an electron donor(s) of Photosystem II (PSII), and oxidation of another unknown intermediate HY to Y would correspond to today's oxidation of electron donor(s) of Photosystem I (PSI). The nature of the HY and HZ, as well as another intermediate labeled as "X" was, of course, unknown then. However, after a decade, Rabinowitch (1956) implied that a cytochrome (Cyt) might be Y, the electron carrier, mentioned above. The reason given for this idea was the experimental observations of reversible light-induced oxidation of Cyt *f*, by Louis N. M. Duysens (1954) (b.1921–d.2015; see Govindjee and Pulles 2016), and by Henrik G. Lundegårdh (1954) (b.1888–d.1969; see Larkum 2003).

Based on the available data, Rabinowitch (1956, see paragraph 2 on page 1862) suggested that a photochemical transfer of electrons from a reduced Cyt to an organic acceptor [perhaps via DPN (now NAD), or TPN (now NADP)] must occur. He added that a transfer of hydrogen [or electrons] from H_2O to the oxidized Cyt would require



Rabinowitch (1945)

Fig. 1 A two-photoreaction scheme for photosynthesis presented by Rabinowitch (1945), which in part, was based on some thoughts by Franck and Herzfeld (1941). Govindjee et al. (2012) suggested that HZ to Y system is equivalent to photosystem II, whereas HY to X system is equivalent to photosystem I, the enzyme E_C is equivalent to OEC (Oxygen Evolving Complex; Mn_4CaO_5 cluster), and E_B to Calvin-Benson cycle enzymes. Also see discussion in Govindjee and Björn (2015)

another photochemical reaction, and then he wrote that the quantum requirement of [all the four] hydrogen [electron] transfer reaction(s) as a whole would be (at least) eight, “since two quanta will be needed to transfer each of the four required H atoms (or electrons), first from water to the cytochrome, and then from the Cyt to the final acceptor.” This has been clearly recognized by Duysens (1989; see page 74, paragraph 2, left column). Thus, we see that the very basis of the series scheme of photosynthesis was already born in 1956!

Experimental discovery of the two light reactions and two pigment systems and the Z-scheme of photosynthesis (1957–1960)

The very first key experiment that brought before us the concept of the two light reaction two-pigment system was the discovery of the *Emerson Enhancement Effect* in oxygenic photosynthesis (Emerson et al. 1957; Emerson and Chalmers 1958; Emerson and Rabinowitch 1960; also see; Govindjee 2004). Robert Emerson suggested that one of the light reactions was run by chlorophyll (Chl) *a* (his long-wave system), but the other by one of the auxiliary pigments (e.g., Chl *b*) (Emerson and Chalmers 1958). This ran against the known fact from Duysens (1952) that excitation energy transfer from Chl *b* to Chl *a* is 100%. Further, Govindjee and Rabinowitch (1960) and R. Govindjee et al. (1960) showed that both photosystems were run by Chl *a*, but of different spectral forms. An important point was that C. Stacy French (1907–1995; see Govindjee and Fork 2006; cf. French 1979), and Cedersstrand et al. (1966) had been able to see these spectral

forms of Chl *a* through absorption spectroscopy (also see Krasnovsky 1992). In view of the fact that Emerson's experiments were made with manometry that cannot distinguish between light-induced changes in oxygen evolution (photosynthesis) and oxygen uptake (respiration), it was important to prove that the effect was not in respiration. This was achieved by inhibiting respiration by parabenzoquinone, which then also acted as electron acceptor instead of CO₂; using this method, R. Govindjee et al. (1960) showed that the two-light effect was not in respiration, but in the Hill reaction (the light reaction phase of photosynthesis). This laid to rest the idea by Lawrence Rogers Blinks (1957, 1959) (b.1900–d.1989; see Abbott and Smith 2010) that the two-light effect may have been in respiration. In addition to the two light effect in photosynthesis, an “antiparallel” two light effect was discovered in Chl *a* fluorescence, as expected (Govindjee et al. 1960), studied soon thereafter, by e.g., Duysens and Sweers (1963). For further discussion on what information Chl *a* fluorescence has provided to the two light reaction two pigment system concept, see chapters in: Papa-georgiou and Govindjee (2004) and in Demmig-Adams et al. (2014). Further, the Emerson enhancement effect was confirmed, by mass spectrometry, to be in photosynthesis (Govindjee et al. 1963), as well as by the existence of Emerson enhancement effect in NADP reduction in chloroplasts (Govindjee et al. 1964). *The scene was set soon thereafter* (see Nickelsen 2015).

A major event at that time was the publication of the famous highly cited and recognized paper by Hill and Bendall (1960) [see Bendall (1994) for an article on Robert (Robin) Hill (1899–1991)]. Based on thermodynamic arguments, a theoretical Z-scheme was published, where, one light reaction (now I) oxidized a Cyt *f* and another light reaction (now II) reduced Cyt *b*₆, and ATP was produced due to energy available from the downhill electron transfer from reduced Cyt *b*₆ to Cyt *f* (see Fig. 2). Although Hill and Bendall (1960) did not relate this scheme to Emerson's discovery and to the concepts of Eugene Rabinowitch, and although the Cyt *b*₆ in this scheme is not the one involved as suggested, this scheme was powerful in its impact; it tied all things together. We note that Robin Hill was indeed a major discoverer; see two excellent articles on him as well as on the Z-Scheme by Walker (2002a, b). We, however, recognize that the idea of two light reaction and two pigment system was “in the air” so to say, and discussed in several publications (see above), and in Kok (1959); and in papers presented, during March 28–31, 1960, by Rabinowitch and Govindjee (pp. 378–586); B. Kok and G. Hoch (pp. 397–416); R. Hill and W.D. Bonner (424–435); C.S. French (pp. 447–471); and D.I. Arnon (pp. 489–565)] at a symposium on “Light and Life” held at the Johns Hopkins

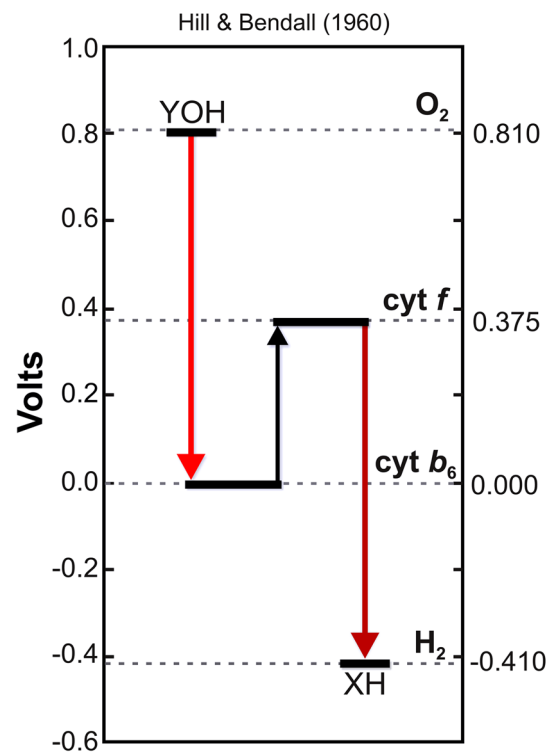


Fig. 2 The two light reaction scheme of Hill and Bendall (1960). The figure was drawn with the water/oxygen system at the top, rather than the other way around, as is done currently to show that the light reaction is an uphill process. In the light reaction related to current photosystem II, a reductant “Y” reduces oxidized Cytochrome (Cyt) *b*₆ becoming YOH, and reducing Cyt *b*₆ (we now know that Cyt *b*₆ does not play this role). In the other light reaction, Cyt *f* reduces an intermediate “X” to “XH”. What was really new in this scheme was that electron flow from reduced Cyt *b*₆ to oxidized Cyt *f* would be a thermo-chemically downhill process, and, thus, having the potential of making ATP just as happens in mitochondria. The diagram shown here was redrawn from the original Fig. 4 of Hill and Bendall (1960)

University; see McElroy and Glass (1961). Hill (1965) presented a detailed and thorough review on the electron transport chain, especially on the Z-scheme of photosynthesis, but, unfortunately, he missed citing Emerson's and Rabinowitch's key papers, on the concept of two light reactions and two pigment systems, cited here.

Evolution of the many versions of the Z-scheme: 1961—the present

The most crucial experiment that clinched the series scheme of photosynthesis was that of Louis N. M. Duysens et al. (1961) who showed an antagonistic effect of light I and II on the redox state of Cyt *f*; Duysens and coworkers first added “red light” to a suspension of red alga *Porphyridium*, and thus, it was *light 1*: it oxidized Cyt *f*; however, when they added the second beam of light, *light 2*, oxidized

Cyt *f* was reduced! Thus, the names *Light I*, *Light II*, *Pigment System I*, *Pigment System II*, *Light Reaction I*, *Light Reaction II*, *Photosystem I (PSI)* and *Photosystem II (PSII)* came into being (also see Duysens and Ames 1962; Duysens 1989; Govindjee and Pulles 2016)! A lesser-known, but an important, experiment that had preceded this experiment was that of Bessel Kok (1918–1979; see Kok 1959; Myers 1987), where he observed that in a cyanobacterium far-red light oxidized the reaction center P700 that he had discovered earlier (Kok 1956, 1957), and a shorter wave light reduced this oxidized P700. Both the photosystems (I and II) as well as both the reaction centers (P700 in PSI; and P680 in PSII) contain Chl *a*, but these different Chl *a*'s have different properties and functions since they are bound differently to different proteins (see Björn et al. 2009 for a perspective).

In this historical minireview, we are not going to discuss the details of the Z-scheme except that we mention the discovery of the reaction center of PSII, P680, by the research group of Horst T. Witt (1922–2007; see Döring et al. 1968; Govindjee et al. 1970; also see; Witt 2004); for early speculations, see Krey and Govindjee (1964) and Rabinowitch and Govindjee (1965), and the discovery of pheophytin function as one of the “primary” electron acceptors of PSII (Klimov et al. 1977, and reviews by; Klimov 2003; and, Mamedov et al. 2015). Today, there are many versions of the Z-scheme, but we show here just a few: a 1965 scheme by Rabinowitch and Govindjee (Fig. 3); another (Fig. 4) by Blankenship and Prince (1985), where they had, in our opinion, correctly included P680, P680*, P700 and P700* as participants in the scheme, and had discussed why excited singlet state potentials of the chlorophylls should be included. We note that even at that time some textbooks (see e.g. Parson 1983) and research papers (Prince et al. 1976; Parson and Monger 1976) had this information in schemes on “anoxygenic photosynthesis”, but more relevant to this paper, on oxygenic photosynthesis were the schemes of Witt (1971), and that of Govindjee and van Rensen (1978). Figure 5 is a simplified scheme of Govindjee and van Rensen (1978) and Fig. 6 is a detailed scheme in the form of “Z-” (modified from Demeter and Govindjee 1989, which was based partly on Fig. 11 in Govindjee and Govindjee 1975). See Supplementary Material for additional figures.

We refer the readers to Mohapatra and Singh (2015) and Jaiswal et al. (2016) to see how the Z-scheme can be taught to students, through drama, outdoors, and indoors, respectively. In addition, for school children, a simple and fun video (DVD), with rap music is available (see: <http://www.biologymusicvideos.com/biol-o-gee-rap-photosynthesis.html>).

Further, we mention four things of special interest to all of us: (1) In addition to linear electron flow from water to

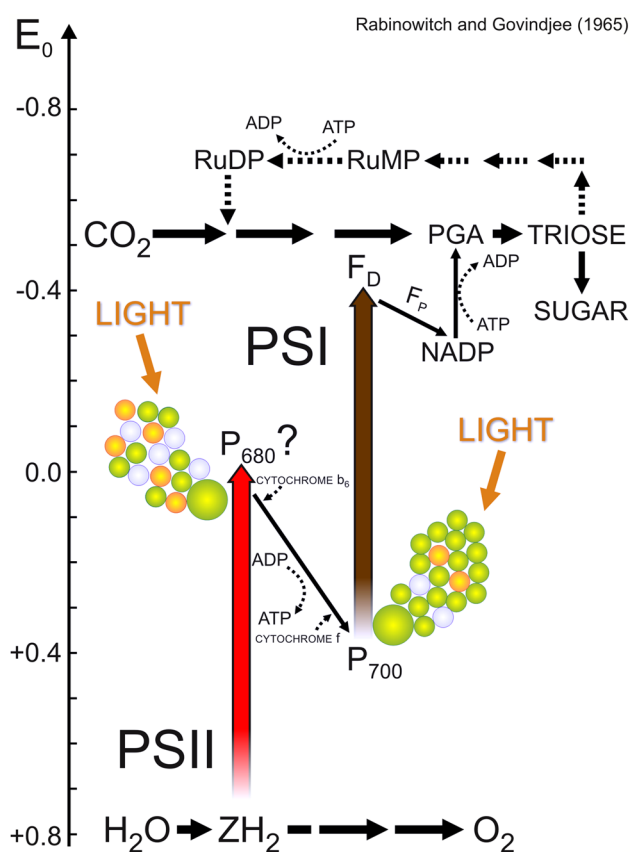
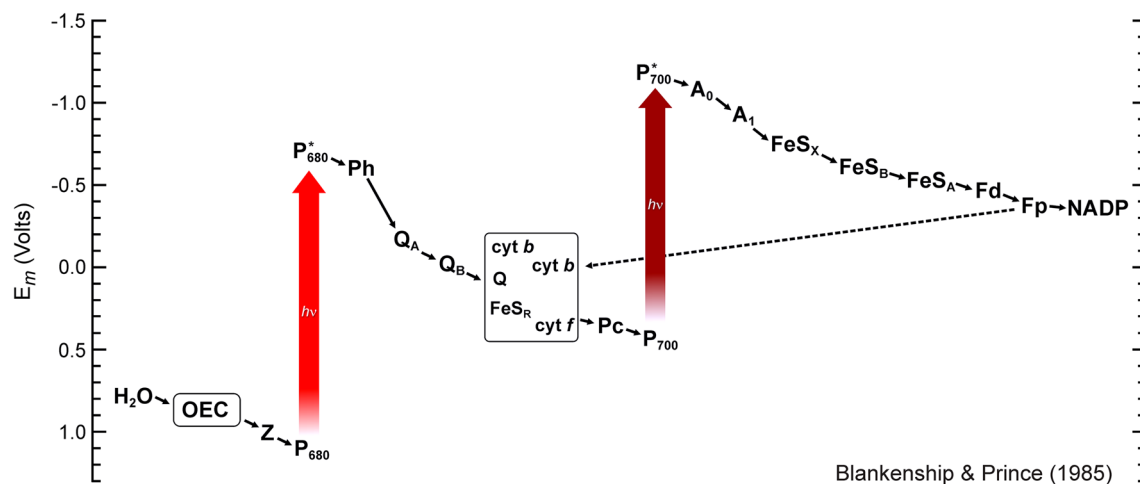


Fig. 3 A 1964–1965 scheme of two light reactions (I and II) and two photosystems (PSI and PSII), which included basic reactions of Calvin-Benson cycle, and of the possible existence of “Trap II”, listed as “P680?” before its discovery by Döring et al. (1968) in Horst Witt’s lab in Berlin (also see Govindjee et al. 1970). ZH_2 is the electron donor to the reaction center (or “Trap II”), whereas P700 is “Trap I”, discovered by Kok (1956, 1957). In retrospect, P680, and the PSII antenna should have been placed close to water. F_D stands for ferredoxin, F_P for a flavoprotein, NADP for nicotinamide dinucleotide phosphate, PGA for phosphoglyceric acid, RuMP and RuDP (now RuBP) for ribulose mono and di (or bi) phosphate. This diagram was made by Govindjee (1964, unpublished; see Krey and Govindjee 1964), and drawn by Natalie H. Davis, an artist in the then Department of Botany, University of Illinois at Urbana-Champaign (UIUC). It was first published by Rabinowitch and Govindjee (1965). [A photograph of Wolfgang Junge with the original 1964–1965 diagram pointing to P680 is in the Supplementary Material (also see a pdf on Junge at <http://www.life.illinois.edu/govindjee/honorsfrom.html>)]

NADP^+ , there exists a “Q” cycle involving Cyt *b₆f* complex, which brings extra protons to the thylakoid lumen, providing greater amount of proton motive force (pmf) to make more ATP molecules per electron transferred (see e.g., Crofts 2004; Dumas et al. 2016, and chapters in; Cramer and Kallas 2016). (2) In almost all plants, there also exists a cyclic electron flow around PSI leading to ATP formation (see e.g., Finazzi and Johnson 2016; Shikanai 2016). (3) Oxygenic photosynthesis uniquely requires bicarbonate ions (hydrogen carbonate) bound on non-heme



Blankenship & Prince (1985)

Fig. 4 A colored (redrawn) version of the Z-scheme from Blankenship and Prince (1985). The redox carriers were placed according to the accepted midpoint redox potentials (pH 7). *OEC*: oxygen evolving complex, *Z*: electron donor to photosystem II (PSII), *P*₆₈₀: reaction center chlorophyll *a* of PSII, *Ph*: pheophytin, *Q*: quinone, *Cyt*: cytochrome, *FeS_R*: Reiske iron-sulfur protein, *Pc*: plastocyanin, *P*₇₀₀: reaction center Chl *a* of photosystem I (PSI), *A*₀ and *A*₁: primary elec-

tron acceptors of PSI, *FeS_X*, *FeS_B*, and *FeS_A*: bound iron-sulfur protein acceptors of PSI, *Fd*: soluble ferredoxin, *Fp*: flavoprotein (ferredoxin-NADP reductase). In this diagram, electron transfer through the redox components of plastoquinone pool and Cyt *b*₆*f* complex is not shown. The dashed line indicates cyclic electron transfer around PSI. [Note that in the current literature some of the redox components of the Z-scheme use different abbreviations.]

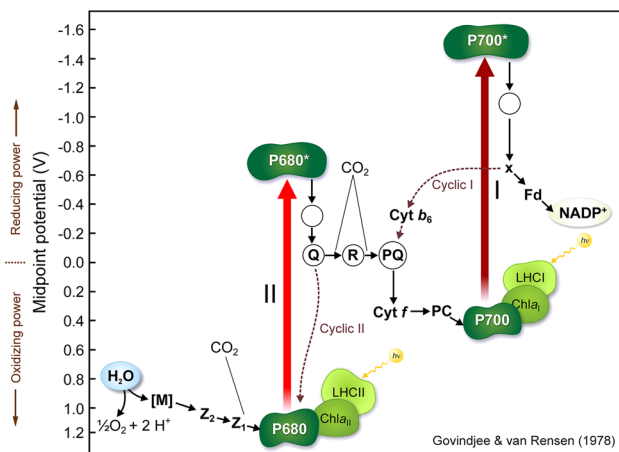


Fig. 5 A simplified (redrawn) and colored version of the Z-scheme from Govindjee and van Rensen (1978). Following Witt (1971), this was the first detailed “visualization” of the excited singlet states of reaction center chlorophylls of photosystem II (PSII) and photosystem I (PSI) (P680 and P700) on top of excitation arrows (light red for PSII and dark red for PSI). Note that the knowledge about photosynthesis at that time was not complete for locating all (presently known) redox cofactors of electron transfer chain into this scheme. For original abbreviations of all shown components, see the detailed legend to Fig. 1 in Govindjee and van Rensen (1978); also see the legend of Fig. 6

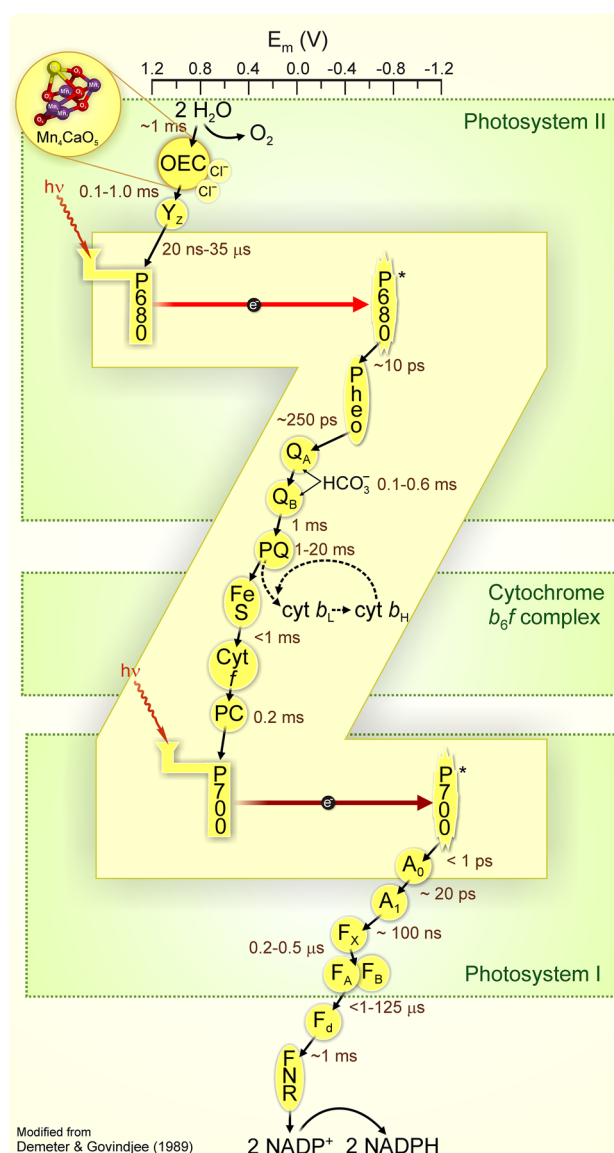
iron between the first and the second plastoquinone electron acceptors *Q_A* and *Q_B*, and this bicarbonate is essential for electron and proton transfer at the *Q_B* site (Wydrzynski and Govindjee 1975; Eaton-Rye and Govindjee 1988; Brinkert et al. 2016; and a review by; Shevela et al. 2012).

Moreover, bicarbonate is also known to have effects on the water-splitting side of PSII (Stemler et al. 1974); unbound bicarbonate ions, on this side, may act as proton acceptors (see e.g., Ananyev et al. 2005; Shutova et al. 2008; Shevela et al. 2013; Koroidov et al. 2014), or participate in photoassembly (Allakhverdiev et al. 1997; Baranov et al. 2004; Dasgupta et al. 2008), or stabilize the water-oxidizing complex (Klimov et al. 1997, 2003), but not to act as a substrate for oxygen evolution (see e.g., Clausen et al. 2005; Hillier et al. 2006). (4) It is of paramount importance to realize that the very first steps that start the photochemistry in oxygenic photosynthesis is the primary charge separation at the reaction center I (P700) and the reaction center II (P680), and that these charge separation events occur within picosecond (10^{-12} s) time scale (see e.g., early papers by Wasielewski et al. 1987, 1989; and reviews by; Mamedov et al. 2015; Milanovsky et al. 2014; Nadochenko et al. 2014).

Shevela and Govindjee (2016) have produced a detailed poster of oxygenic photosynthesis; it is available free: at http://www.life.illinois.edu/govindjee/Electronic%20Publications/2016_Poster-Shevela_Govindjee.pdf. It includes the oxygen-evolving manganese clock (see e.g., Joliet and Kok 1975); the ATP synthesis clock (see e.g., Mitchell 1961; Abrahams et al. 1994; Boyer 1997; Jagendorf 2002; Junge 2004); the two-electron gate for plastoquinone reduction (see e.g., chapters in Wydrzynski and Satoh 2005); and the carbon reduction cycle, the Calvin-Benson cycle (see e.g., Benson 2002; Bassham 2003).

We end this perspective with the news that tremendous progress has now been made in understanding both the

Fig. 6 The Z scheme of electron transport in photosynthesis. The electron carriers are placed horizontally according to their midpoint redox potentials at pH 7.0 (E_m 7). A slightly modified version of the original 1989 figure legend follows. Electron flow is initiated when a photon or exciton reaches the reaction center Chl *a* P680 (in PS II) and P700 (in PS I) (see $h\nu$ going into the two funnels). P680* and P700* (see distorted ovals) indicate the first singlet excited states of P680 and P700. The *first* reaction of P680* is the conversion of excitonic energy into chemical energy: charge separation, i.e., the formation of the cation P680⁺ and the anion (Pheo⁻) within ~10 ps (a chlorophyll before pheophytin has been left out). [For information on pheophytin, see Allakhverdiev et al. 2010.] The first reaction of P700*, the charge separation into P700⁺A₀⁻, may need <1 ps. Here, A₀ is a special Chl *a* molecule. The P680* recovers its lost electron from Y_Z, tyrosine-161 of the D1 polypeptide of PSII. The positive charge on Y_Z is then transferred to the charge accumulator Mn₄CaO₅ cluster, or the oxygen-evolving complex (OEC). Four positive charges must accumulate before an O₂ molecule is evolved. The Pheo⁻ delivers the “extra” electron to a primary (plastoquinone) electron acceptor, Q_A located on the D2 polypeptide of PSII; Q_A⁻ delivers its electron to a secondary (plastoquinone) electron acceptor O_B⁻ located on the D1 polypeptide of PSII. After reduction to plastoquinol, i.e. after two turnover of the P680, Q_B(H₂) exchanges with a mobile plastoquinone (PQ) molecule. Bicarbonate ions (HCO₃⁻) are now known to be involved in the Q_A-Fe-Q_B region, where Fe is a non-heme iron atom between the two plastoquinones. Plastoquinol (PQH₂) delivers one electron to the Rieske iron-sulfur protein (FeS), and the other to a Cytb_L. The electron on reduced FeS reduces Cyt *f*, and the one on Cyt b_L is transferred to Cyt b_H, returning back in a cyclic process (called the Q-cycle). Reduced Cyt *f* delivers its extra electron to a copper protein, plastocyanin (PC), which delivers the electron to P700⁺ (produced in the primary PSI reaction). On the other hand, A₀ passes its electron to A₁ (a phytylquinone molecule). The rest of the electron carriers are: F_x, F_A, and F_B (iron sulfur clusters), Fd (ferredoxin) and FNR (ferredoxin-NADP⁺ reductase). The diagram shows either measured or estimated times of the various reactions in the Z-scheme, except for the production of P680* and P700* that occur in femtosecond time scale. The bottleneck reaction is of the order of 5 ms and it involves the total time involved in the exchange of Q_B(H₂) with PQ; and the diffusion of PQH₂ to the Cyt b₆f complex, and the reoxidation time of PQH₂ (modified from Fig. 2 in Demeter and Govindjee 1989)



photosystems at atomic level resolution, and it is obvious to us that we are now very close to understanding both the physics and chemistry of excitation energy transfer as well as photochemical reactions in both photosystems since atomic level crystal structures of both PSI and PSII are now available. Even before this, Crofts et al. (1987) and Xiong et al. (1996), among others, had obtained atomic level PSII structure, based on homology modeling with the bacterial reaction centers [see Xiong et al. (1998) for a detailed review]. Now, high resolution (1.9–3.5 Å) structures of PSII are available, not only for thermophilic cyanobacteria (Umena et al. 2011; Shen 2015; Suga et al. 2015; Young et al. 2016), but also for red algae at 2.76 Å resolution (Ago et al. 2016), and for higher plants at 3.2 Å resolution (Wei et al. 2016) (Fig. 7). As a starting point, readers are referred to the following publications, particularly on PSII: Amunts et al. (2007); Barber (2016); Cox et al. (2014); Mukherjee et al. (2012), Najafpour

et al. (2012, 2013); Nelson and Junge (2015); Qin et al. (2015); Vinyard et al. (2013); and Zhang et al. (2015). *The future of our knowledge about photosynthesis looks very bright.* This paper is a good example of how fast our knowledge can evolve just within several decades. For information on just a few selected leaders in the field, see presentations at: <http://www.life.illinois.edu/govindjee/honorsfrom.html>.

Readers are encouraged to consult Walker (1992a, b); and books by Rabinowitch and Govindjee (1969), Ke (2001), Blankenship (2014), and in the near future a book by Björn et al. (forthcoming). For further discussion on different aspects of photosynthesis, see various volumes in *Advances in Photosynthesis and Respiration* (<http://www.springer.com/series/5599>), including those edited by Ort and Yocum (1996; on Oxygenic Photosynthesis),

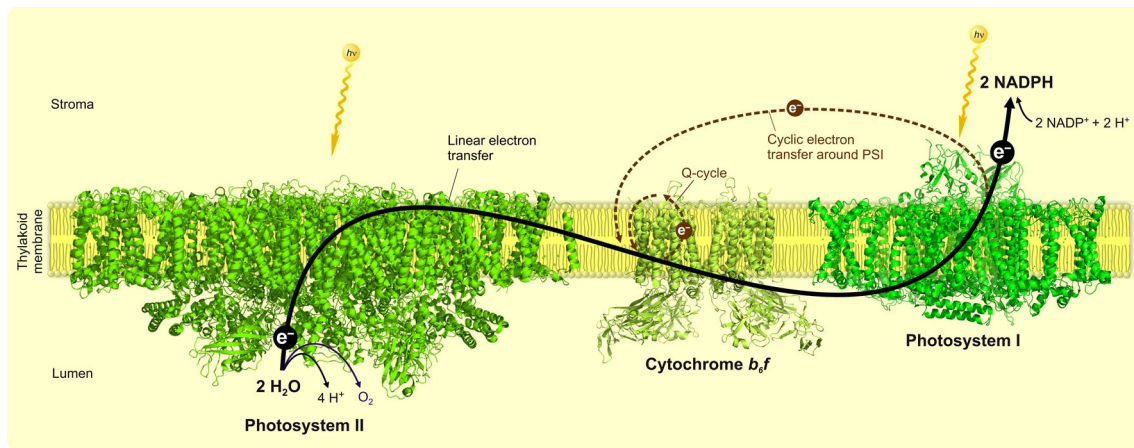


Fig. 7 Overall side-on view of the three major multiprotein photosynthetic complexes (PSII, Cyt *b₆f*, and PSI), located in the thylakoid membrane; they contain (or bind) the redox components (not shown) required for linear electron flow from H₂O to NADP⁺ (**bold black arrow**). The higher plant (spinach) dimeric PSII-LHCII supercomplex was generated using coordinates of cryo-EM structure at 3.2 Å depos-

ited at Protein Data Bank (PDB) as ID 3JCU (Wei et al. 2016). The dimeric Cyt *b₆f* complex of the thermophilic cyanobacterium *Mastigocladus laminosus* was generated from a 3.0 Å crystal structure deposited at PDB as ID 1VF5 (Kurusu et al. 2003). The higher plant (spinach) PSI-LHCI supercomplex at 2.8 Å resolution was produced employing coordinates from PDB, using ID 4Y28 (Mazor et al. 2015)

Wydrzynski and Satoh (2005; on Photosystem II), Govindjee et al. (2005; on Discoveries in Photosynthesis), Golbeck (2006; on Photosystem I), Eaton-Rye et al. (2012; on Overviews on Photosynthesis) and Cramer and Kallas (2016; on Cytochromes).

Acknowledgements We are highly indebted to Robert Blankenship, Wim Vermaas, John Raven, and four reviewers of this manuscript, who helped us improve the presentation of this historical educational paper. We thank P. Lester Dutton for looking at our Fig. 6 and for bringing to our attention reviews on the primary photochemistry of photosynthesis. Govindjee thanks Rajni Govindjee for her support during the preparation of this paper; he is grateful to the excellent staff of the offices of Information Technology (Life Sciences), Plant Biology, Biochemistry, and Biophysics & Quantitative Biology of the University of Illinois at Urbana-Champaign for their cooperation and help in all what he does. The proofs were corrected on January 14, 2017, when Govindjee was visiting the laboratories of Ashwani Pareek and Baishnab Tripathy at Jawaharlal Nehru University, New Delhi, India.

Appendix

The following example shows that science is a self-correcting enterprise, no matter who the authors are, and that even the top scientists make mistakes. James Franck, together with Gustav Ludwig Hertz, received the 1925 Nobel Prize in Physics in 1926 for “for their discovery of the laws governing the impact of an electron upon an atom”. Later, he became known for the “Franck–Condon Principle”, which states that upon light absorption, a molecule goes into an excited state, but in a higher vibrational

state. (See Rice and Jortner (2010) for all the major contributions, and life, of Franck.) Franck contributed extensively to photosynthesis (see Rosenberg 2004; also see Franck and Rosenberg 1964). Unfortunately, some of his thoughts, which may have been physically sound, turned out to be incorrect. Examples are: instead of using realistic 3-dimensional structure of the “antenna”, Franck and Teller (1938) calculated excitation energy transfer as if the pigments were located in one dimension; with these results, they challenged the concept of “photosynthetic unit”, but when two-dimensional and multidimensional approaches were used, their conclusions could not be accepted (see e.g., Bay and Pearlstein 1963; Robinson 1967). In the same manner, explanation by Franck (1958) of the “red drop” (Emerson and Lewis 1943) and the Emerson Enhancement Effect (Emerson et al. 1957), by double excitation (“up-conversion”) of the same chlorophyll *a* molecules was also incorrect.

References

- Abbott IA, Smith CM (2010) Lawrence Rogers Blinks (1900–1989). Biographical Memoirs of the National Academy of Sciences, USA, pp. 3–19
- Abrahams JP, Leslie AG, Lutter R, Walker JE (1994) Structure at 2.8 Å resolution of F1-ATPase from bovine heart mitochondria. *Nature* 370:621–628
- Ago H, Adachi H, Umena Y, Tashiro T, Kawakami K, Kamiya N, Tian L, Han G, Kuang T, Liu Z, Wang F, Zou H, Enami I, Miyano M, Shen JR (2016) Novel features of eukaryotic photosystem II

- revealed by its crystal structure analysis from a red alga. *J Biol Chem* 291:5676–5687
- Allakhverdiev S, Yruela I, Picorel R, Klimov V (1997) Bicarbonate is an essential constituent of the water-oxidizing complex of photosystem II. *Proc Natl Acad Sci USA* 94: 5050–5054
- Allakhverdiev SI, Tomo T, Shimada Y, Kindo H, Nagao R, Klimov VV, Mimuro M (2010) Redox potential of pheophytin *a* in photosystem II of two cyanobacteria having the different special pair chlorophylls. *Proc Natl Acad Sci USA* 107:3924–3929
- Amunts A, Drory O, Nelson N (2007) The structure of a plant photosystem I supercomplex at 3.4 Å resolution. *Nature* 447:56–63
- Ananyev G, Nguyen T, Putnam-Evans C, Dismukes GC (2005) Mutagenesis of CP43-arginine-357 to serine reveals new evidence for (bi)carbonate functioning in the water oxidizing complex of photosystem II. *Photochem Photobiol Sci* 4:991–998
- Bannister TT (1972) The careers and contributions of Eugene Rabinowitch. *Biophys J* 12:707–718
- Baranov SV, Tyryshkin AM, Katz D, Dismukes GC, Ananyev GM, Klimov VV (2004) Bicarbonate is a native cofactor for assembly of the manganese cluster of the photosynthetic water oxidizing complex. Kinetics of reconstitution of O₂ evolution by photoactivation. *Biochemistry* 43:2070–2079
- Barber J (2016) Mn₄Ca cluster of photosynthetic oxygen-evolving center: structure, function and evolution. *Biochemistry* 55:5901–5906. doi:10.1021/acs.biochem.6b00794
- Bassham JA (2003) Mapping the carbon reduction cycle: a personal retrospective. *Photosynth Res* 76:35–52
- Bay Z, Pearlstein RM (1963) A theory of energy transfer in the photosynthetic unit. *Proc Natl Acad Sci USA* 50:1071–1078
- Bendall DS (1994) Robert Hill (1899–1991). *Biogr Memoirs Fellows R Soc* 40:142–170
- Benson AA (2002) Following the path of carbon in photosynthesis: a personal story. *Photosynth Res* 73:29–49
- Björn LO, Papageorgiou GC, Blankenship R, Govindjee (2009) A viewpoint: why chlorophyll *a*? *Photosynth Res* 99:85–98
- Björn LO, Shevela D, Govindjee (forthcoming) *Photosynthesis*. World Scientific Publishing Co, Singapore
- Blankenship RE (2014) *Molecular mechanisms of photosynthesis*. Blackwell/Wiley, Malden
- Blankenship RE, Prince RC (1985) Excited state redox potentials and the Z scheme of photosynthesis. *Trends Biochem Sci* 10:382–383
- Blinks LR (1957) Chromatic transient in photosynthesis of red algae. In: Gaffron H, Brown AH, French CS, Livingston R, Rabinowitch EI, Strehler B, Tolbert NE (eds) *Research in photosynthesis*. Interscience Publishers, New York, pp 444–449
- Blinks LR (1959) Chromatic transients in the photosynthesis of a green alga. *Plant Physiol* 34:200–203
- Boyer PD (1997) The ATP synthase: a splendid molecular machine. *Annu Rev Biochem* 66:717–749
- Brinkert K, De Causmaecker S, Krieger-Liszkay A, Fantuzzi A, Rutherford AW (2016) Bicarbonate-induced redox tuning in Photosystem II for regulation and protection. *Proc Natl Acad Sci USA* 113:12144–12149
- Cederstrand C, Rabinowitch E, Govindjee (1966) Analysis of the red absorption band of chlorophyll *a* in vivo. *Biochim Biophys Acta* 126:1–12
- Clausen J, Beckman K, Junge W, Messinger J (2005) Evidence that bicarbonate is not the substrate in photosynthetic oxygen evolution. *Plant Physiol* 139:1444–1450
- Cox N, Retegan M, Neese F, Pantazis DA, Boussac A, Lubitz W (2014) Electronic structure of the oxygen-evolving complex in photosystem II prior to O–O bond formation. *Science* 345:804–808
- Cramer WA, Kallas T (eds) (2016) *Cytochrome complexes: evolution, structures, energy transduction, and signaling*. Advances in photosynthesis and respiration, vol 41. Springer, Dordrecht
- Crofts AR (2004) The Q-cycle: a personal perspective. *Photosynth Res* 80:223–243
- Crofts AR, Robinson HH, Andrews K, Van Doren S, Berry E (1987) Catalytic sites for reduction and oxidation of quinones. In: Papa S, Chance B, Ernster L (eds) *Cytochrome systems: molecular biology and bioenergetics*. Plenum Press, NY, pp 617–624
- Dasgupta J, Ananyev GM, Dismukes GC (2008) Photoassembly of the water-oxidizing complex in photosystem II. *Coord Chem Rev* 252:347–360
- Demeter S, Govindjee (1989) Thermoluminescence in plants. *Physiologia Plant* 75:121–130
- Demmig-Adams B, Garab G, Adams WW III, Govindjee (eds) (2014) *Non-photochemical quenching and energy dissipation in plants, algae and cyanobacteria*. Advances in photosynthesis and respiration, vol 40. Springer, Dordrecht
- Döring G, Bailey JL, Kreutz W, Witt HT (1968) The active chlorophyll-*a*-II in light reaction II of photosynthesis. *Naturwissenschaften* 55:220–221
- Dumas L, Chazaux M, Peltier G, Johnson X, Alric J (2016) Cytochrome *b₆f* function and localization, phosphorylation state of thylakoid membrane proteins and consequences on cyclic electron flow. *Photosynth Res* 129:307–320
- Duysens LNM (1952) Transfer of excitation energy in photosynthesis. Doctoral Thesis. State University, Utrecht
- Duysens LNM (1954) Role of cytochrome and pyridine nucleotide in algal photosynthesis. *Science* 121:210–211
- Duysens LNM (1989) The discovery of the two photosynthetic systems: a personal account. *Photosynth Res* 21:61–80
- Duysens LNM, Ames J (1962) Function and identification of two photochemical systems in photosynthesis. *Biochim Biophys Acta* 64:243–260
- Duysens LNM, Sweers HE (1963) Mechanism of two photochemical reactions in algae as studied by means of fluorescence. In: Japanese Society of Plant Physiologists (eds), *Studies on microalgae and photosynthetic Bacteria*. University of Tokyo Press: Tokyo, pp 353–372
- Duysens LNM, Ames J, Kamp BM (1961) Two photochemical systems in photosynthesis. *Nature* 190:510–511
- Eaton-Rye JJ, Govindjee (1988) Electron transfer through the quinone acceptor complex of Photosystem II after one or two actinic flashes in bicarbonate-depleted spinach thylakoid membranes. *Biochim Biophys Acta* 935:248–257
- Eaton-Rye JJ, Tripathy BC, Sharkey TD (eds) (2012) *Photosynthesis: plastid biology, energy conversion and carbon assimilation*. Advances in photosynthesis and respiration, vol 34. Springer, Dordrecht
- Emerson R, Chalmers RV (1958) Speculations concerning the function and phylogenetic significance of the accessory pigments of algae. *Phycol Soc News Bull* 11:51–56
- Emerson R, Lewis CM (1943) The dependence of the quantum yield of *Chlorella* photosynthesis on wavelength of light. *Am J Bot* 30:165–178
- Emerson R, Rabinowitch E (1960) Red drop and role of auxiliary pigments in photosynthesis. *Plant Physiol* 35:477–485
- Emerson R, Chalmers RV, Cederstrand CN (1957) Some factors influencing the long wave limit of photosynthesis. *Proc Natl Acad Sci USA* 43:133–143
- Finazzi G, Johnson GN (2016) Cyclic electron flow: facts and hypotheses. *Photosynth Res* 129:227–230
- Franck J (1958) Remarks on the long-wavelength limits of photosynthesis and chlorophyll fluorescence. *Proc Natl Acad Sci USA* 44:941–948

- Franck J, Herzfeld KF (1941) Contribution to a theory of photosynthesis. *J Phys Chem* 45:978–1025
- Franck J, Rosenberg JL (1964) A theory of light utilization in plant photosynthesis. *J Theor Biol* 7:276–301
- Franck J, Teller E (1938) Migration and photochemical action of excitation energy in crystals. *J Chem Phys* 6:861–872
- French CS (1979) Fifty years of photosynthesis. *Annu Rev Plant Physiol* 30:1–36
- Golbeck JH (ed) (2006) Photosystem I: the light-driven plastocyanin: ferredoxin oxidoreductase. *Advances in photosynthesis and respiration*, vol 24. Springer, Dordrecht
- Govindjee (2004) Robert Emerson, Eugene Rabinowitch: understanding photosynthesis. In: Lillian Hoddeson (ed). “No boundaries: University of Illinois Vignettes”, Chap. 12, pp. 181–194. University of Illinois Press, Urbana and Chicago
- Govindjee, Björn LO (2015) Dissecting oxygenic photosynthesis: the evolution of the “Z”-scheme for Thylakoid reactions. In: Itoh S, Mohanty S, Guruprasad KN (eds) *Photosynthesis: basics to applications*. I. K. International Publishing House Pvt. Ltd., New Delhi, pp 1–27
- Govindjee, Fork DC (2006) Charles Stacy French (1907–1995). *biographical memoirs*, vol 88. National Academy of Sciences, Washington, pp 2–29
- Govindjee, Govindjee R (1975) Introduction to photosynthesis. In: Govindjee (ed) *Bioenergetics of photosynthesis*. Academic Press, New York, pp 2–50
- Govindjee, Pulles MPJ (2016) Louis Nico Marie Duysens (March 15, 1921–September 8, 2015): A leading biophysicist of the 20th century. *Photosynth Res* 128:223–234
- Govindjee, Rabinowitch E (1960) Two forms of chlorophyll *a* in vivo with distinct photochemical functions. *Science* 132:159–160
- Govindjee, van Rensen JJS (1978) Bicarbonate effects on the electron flow in isolated broken chloroplasts. *Biochim Biophys Acta* 505:183–213
- Govindjee, Ichimura S, Cederstrand C, Rabinowitch E (1960) Effect of combining far-red light with shorter wave light in the excitation of fluorescence in *Chlorella*. *Arch Biochem Biophys* 89:322–323
- Govindjee R, Thomas JB, Rabinowitch E (1960) “Second Emerson effect” in the Hill reaction of *Chlorella* cells with quinone as oxidant. *Science* 132:421
- Govindjee, Owens OvH, Hoch G (1963) A mass spectroscopic study of the Emerson enhancement effect. *Biochim Biophys Acta* 75:281–284
- Govindjee R, Govindjee, Hoch G (1964) Emerson enhancement effect in chloroplast reactions. *Plant Physiol* 39:10–14
- Govindjee R, Rabinowitch E, Govindjee (1968) Maximum quantum yield and action spectrum of photosynthesis and fluorescence in *Chlorella*. *Biochim Biophys Acta* 162:539–544
- Govindjee, Döring G, Govindjee R (1970) The active chlorophyll *a* II in suspensions of lyophilized and tris-washed chloroplasts. *Biochim Biophys Acta* 205:303–306
- Govindjee, Beatty JT, Gest H, Allen JF (eds) (2005) *Discoveries in photosynthesis. Advances in photosynthesis and respiration*, vol 20. Springer, Dordrecht
- Govindjee, Björn LO, Nickelsen K (2012) Evolution of the Z-scheme of electron transport in oxygenic photosynthesis. C Lu (ed) *Photosynthesis: research for food, fuel and future—15th International Conference on Photosynthesis, Symposium: Education Session*, University Press, Springer-Verlag GmbH, Zhejiang, pp 835–841
- Hill R (1965) Biochemists’ green mansions: the photosynthetic electron transport chain in plants. *Essays Biochem* 1:121–151
- Hill R, Bendall F (1960) Function of the two cytochrome components of chloroplast: a working hypothesis. *Nature* 186:136–137
- Hill JF, Govindjee (2014) The controversy over the minimum quantum requirement for oxygen evolution. *Photosynth Res* 122:97–112
- Hillier W, MnConnell I, Badger MR, Boussac A, Klimov VV, Dismukes GC, Wydrzynski T (2006) Quantitative assessment of intrinsic carbonic anhydrase activity and the capacity for bicarbonate oxidation in photosystem II. *Biochemistry* 45:2094–2102
- Jagendorf AT (2002) Photophosphorylation and the chemiosmotic perspective. *Photosynth Res* 73:233–241
- Jaiswal S, Bansal M, Roy S, Bharari A, Padhi B (2016) Electron flow from water to NADP⁺ with students acting as molecules in the chain: a Z-scheme drama in the classroom. *Photosynth Res*. doi:10.1007/s1120-016-0317-z
- Joliot P, Kok B (1975) Oxygen evolution in photosynthesis. In: Govindjee (ed) *Bioenergetics of photosynthesis*. Academic Press, NY, pp 388–413
- Junge W (2004) Protons, proteins and ATP. *Photosynth Res* 80:197–221
- Ke B (2001) *Photosynthesis: photobiochemistry and photobiophysics, advances in photosynthesis and respiration*. vol. 10, Springer, Dordrecht
- Klimov VV (2003) Discovery of pheophytin function in the photosynthetic energy conversion as the primary electron acceptor of Photosystem II. *Photosynth Res* 76:247–253
- Klimov VV, Klevanik AV, Shuvalov VA, Krasnovsky AA (1977) Reduction of pheophytin in the primary light reaction of photosystem 2. *FEBS Lett* 82:183–186
- Klimov V, Baranov S, Allakhverdiev S (1997) Bicarbonate protects the donor side of photosystem II against photoinhibition and thermoinactivation. *FEBS Lett* 418:243–246
- Klimov VV, Allakhverdiev SI, Nishiyama Y, Khorobrykh AA, Murata N (2003) Stabilization of the oxygen-evolving complex of photosystem II by bicarbonate and glycinebetaine in thylakoid and subthylakoid preparations. *Funct Plant Biol* 30:797–803
- Kok B (1956) On the reversible absorption change at 705 mμ in photosynthetic organisms. *Biochim Biophys Acta* 22:399–401
- Kok B (1957) Absorption changes induced by the photochemical reaction of photosynthesis. *Nature* 179:583–584
- Kok B (1959) Light-induced absorption changes in photosynthetic organisms. II. A split-beam difference spectrophotometer. *Plant Physiol* 34:184–192
- Koroidov S, Shevela D, Shutova T, Samuelsson G, Messinger J (2014) Mobile hydrogen carbonate acts as proton acceptor in photosynthetic water oxidation. *Proc Natl Acad Sci USA* 111:6299–6304
- Krasnovsky AA (1992) Excited chlorophyll and related problems. *Photosynth Res* 33:177–192
- Krey A, Govindjee (1964) Fluorescence changes in *Porphyridium* exposed to green light of different intensity: a new emission band at 693 nm and its significance to photosynthesis. *Proc Natl Acad Sci USA* 52:1568–1572
- Kurisu G, Zhang H, Smith JL, Cramer WA (2003) Structure of the cytochrome *b₆f* complex of oxygenic photosynthesis: tuning the cavity. *Science* 302:1009–1014
- Larkum AWD (2003) Contributions of Henrik Lundegårdh. *Photosynth Res* 76:105–110
- Lundegårdh H (1954) On the oxidation of cytochrome *f* by light. *Physiol Plant* 7:375–382
- Mamedov M, Govindjee, Nadochenko V, Semenov A (2015) Primary electron transfer processes in photosynthetic reaction centers from oxygenic organisms. *Photosynth Res* 125:51–63
- Mazor Y, Borovikova A, Nelson N (2015) The structure of plant photosystem I supercomplex at 2.8 Å resolution. *eLife* 4:e07433–e07433
- McElroy WD, Glass B (eds) (1961) *A symposium on light and life*. The Johns Hopkins Press, Baltimore

- Milanovsky GE, Ptushenko VV, Cherpanov DA, Semenov AY (2014) Mechanism of primary and secondary ion-radical pair formation in photosystem I complexes. *Biochemistry* 79:221–226
- Mitchell P (1961) Coupling of phosphorylation to electron and hydrogen transfer by a chemiosmotic type of mechanism. *Nature* 191:144–148
- Mohapatra PK, Singh NR (2015) Teaching the Z-scheme of electron transport in photosynthesis: a perspective. *Photosynth Res* 123:105–114
- Mukherjee S, Stull JA, Yano J, Stamatas T, Pringouri K, Stich TA, Abboud KA, Britt RD, Yachandra VK, Christou G (2012) Synthetic model of the asymmetric $[\text{Mn}_3\text{CaO}_4]$ cubane core of the oxygen-evolving complex of Photosystem II. *Proc Natl Acad Sci USA* 109:2257–2262
- Myers J (1987) Bessel Kok (1918–1979) biographical memoirs of the Natl Acad Sci 57:125–148
- Nadtochenko VA, Shelaev IV, Mamedov MD, Shkuropatov AY, Semenov AY, Shuvalov VA (2014) Primary radical ion pairs in photosystem II core complexes. *Biochemistry* 79(3):197–204
- Najafpour MM, Moghaddam AN, Allakhverdiev SI, Govindjee (2012) Biological water oxidation: lessons from Nature. *Biochim Biophys Acta* 1817:1110–1121
- Najafpour MM, Moghaddam AN, Shen J-R, Govindjee (2013) Water oxidation and water-oxidizing complex in cyanobacteria. In: Srivastava AK, Rai AN, Neilan BA (eds) *Stress biology of cyanobacteria*. CRC Publishers, Cleveland, pp. 41–60
- Nelson N, Junge W (2015) Structure and energy transfer in photosystems of oxygenic photosynthesis. *Annu Rev Biochem* 84:659–683
- Nickelsen K (2015) Explaining photosynthesis: models of biochemical mechanisms, 1840–1960. Springer, Dordrecht
- Nickelsen K, Govindjee (2011) The maximum quantum yield controversy. *Otto Warburg and the midwest gang*. Bern Studies in the History and Philosophy of Science, Bern 2011. ISBN: 978-3-9523421-9-0, paperback, p 138
- Ort D, Yocum C (eds) (1996) *Oxygenic photosynthesis: the light reactions advances in photosynthesis and respiration*, vol 4. Springer, Dordrecht
- Papageorgiou G, Govindjee (eds) (2004) *Chlorophyll a fluorescence: a signature of photosynthesis*. *Advances in photosynthesis and respiration*, vol 19. Springer, Dordrecht, p 820
- Parson WW (1983) *Photosynthesis*. In: Zubay G (ed) *Biochemistry*. Addison-Wesley Publishing Company, Boston
- Parson WW, Monger TG (1976) Interrelationships among excited states in bacterial reaction centers. *Brookhaven Symp Biol* 28:195–211
- Prince RG, Leigh JS Jr, Dutton PL (1976) Thermodynamic properties of the reaction center of *Rhodospseudomonas viridis*. In vivo measurement of the reaction center bacteriochlorophyll-primary acceptor intermediary electron carrier. *Biochim Biophys Acta* 440:622–636
- Qin X, Suga M, Kuang T, Shen JR (2015) Photosynthesis. Structural basis for energy transfer pathways in the plant PSI-LHCI supercomplex. *Science* 348:989–995
- Rabinowitch EI (1945) *Photosynthesis*, vol I. Interscience Publishers, New York
- Rabinowitch EI (1956) *Photosynthesis and related processes*, vol II, Part 2. Interscience Publishers, New York
- Rabinowitch E (1961) Robert Emerson (1903–1959). *Biogr Mem Natl Acad Sci USA* 25:112–131
- Rabinowitch E, Govindjee (1965) The role of chlorophyll in photosynthesis. *Sci Am* 213:74–83
- Rabinowitch E, Govindjee (1969) *Photosynthesis*. John Wiley & Sons; copyright by Govindjee. Available free at <http://www.life.illinois.edu/govindjee/g/Books.html>
- Rice SA, Jortner J (2010) James Franck-(1882–1964) *Biographical Memoirs of the National Academy of Sciences*. Washington, DC, pp. 1–28
- Robinson GW (1967) Excitation transfer and trapping in photosynthesis. *Brookhaven Symp Biol* 19:16–48
- Rosenberg JL (2004) The contributions of James Franck to photosynthesis research: a tribute. *Photosynth Res* 80:71–76
- Shen J-R (2015) The structure of photosystem II and the mechanism of water oxidation in photosynthesis. *Annu Rev Plant Biol* 66:23–48
- Shevela D, Eaton-Rye JJ, Shen J-R, Govindjee (2012) Photosystem II and unique role of bicarbonate: a historical perspective. *Biochim Biophys Acta* 1817:1134–1151
- Shevela D, Nöring B, Koroidov S, Shutova T, Samuelsson G, Messinger J (2013) Efficiency of photosynthetic water oxidation at ambient and depleted levels of inorganic carbon. *Photosynth Res* 117:401–412
- Shikanai T (2016) Regulatory network of proton motive force: contribution of cyclic electron transport around photosystem I. *Photosynth Res* 129:253–260
- Shutova T, Kenneweg H, Buchta J, Nikitina J, Terentyev V, Chernyshov S, Andersson B, Allakhverdiev SI, Klimov VV, Dau H, Junge W, Samuelsson G (2008) The photosystem II-associated C_{ah3} in *Chlamydomonas* enhances the O₂ evolution rate by proton removal. *EMBO J* 27:782–791
- Stemler A, Babcock GT, Govindjee (1974) The effect of bicarbonate on photosynthetic oxygen evolution in flashing light in chloroplast fragments. *Proc Natl Acad Sci USA* 71:4679–4683
- Suga M, Akita F, Hirara K, Ueno G, Murakami H, Nakajima Y, Shimizu T, Yamashita K, Ago H, Shen J-R (2015) Native structure of photosystem II at 1.95 Å resolution viewed by femtosecond X-ray pulses. *Nature* 517:99–103
- Tsygankov AA, Allakhverdiev SI, Tomo T, Govindjee (2016) International conference on “Photosynthesis Research for Sustainability-2016”: In honor of Nathan Nelson and Turhan Nejat Veziroglu. *Photosynth Res*. doi:10.1007/s11120-016-0311-5
- Umena Y, Kawakami K, Shen J-R, Kamiya N (2011) Crystal structure of oxygen-evolving photosystem II at a resolution of 1.9 Å. *Nature* 473:55–60
- Vinyard DJ, Ananyev GM, Dismukes GC (2013) Photosystem II: the reaction center of oxygenic photosynthesis. *Annu Rev Biochem* 82:577–606
- Walker DA (1992a) Excited leaves. *Tansley Review No. 36*. *New Phytol* 121:325–345
- Walker DA (1992b) *Energy, plants and man, oxygraphics*. Brighton, East Sussex
- Walker DA (2002a) And whose bright presence—an appreciation of Robert Hill and his reaction. *Photosynth Res* 73(1):51–54
- Walker DA (2002b) The Z-scheme: down hill all the way. *TIPS* 7:183–185
- Wasielowski MR, Fenton JM, Govindjee (1987) The rate of formation of $\text{P700}^+-\text{A}_0^-$ in Photosystem I particles from spinach as measured by picosecond transient absorption spectroscopy. *Photosynth Res* 12:181–190
- Wasielowski MR, Johnson DG, Seibert M, Govindjee (1989) Determination of the primary charge separation rate in isolated Photosystem II reaction centers with 500 femtosecond time resolution. *Proc Natl Acad Sci USA* 86:524–528
- Wei X, Su X, Cao P, Liu X, Chang W, Li M, Zhang X, Liu Z (2016) Structure of spinach photosystem II-LHCII supercomplex at 3.2 Å resolution. *Nature* 534:69–74
- Witt HT (1971) Coupling of quanta, electrons, fields, ions and phosphorylation in the functional membrane of photosynthesis. Results by pulse spectroscopic methods. *Quart Rev Biophys* 4:365–477

- Witt HT (2004) Steps on the way to building blocks, topologies, crystals and X-ray structural analysis of Photosystem I and II of water oxidizing photosynthesis. *Photosynth Res* 80:86–107
- Wydrzynski T, Govindjee (1975) A new site of bicarbonate effect in photosystem II of photosynthesis: evidence from chlorophyll fluorescence transients in spinach chloroplasts. *Biochim Biophys Acta* 387:403–408
- Wydrzynski T, Satoh K (eds) (2005) Photosystem II: the light-driven water:plastoquinone oxidoreductase. *Advances in photosynthesis and respiration*, vol 22. Springer, Dordrecht
- Xiong J, Subramaniam S, Govindjee (1996) Modeling of the D1/D2 proteins and cofactors of the Photosystem II reaction center: implications to herbicide and bicarbonate binding. *Protein Sci* 5:2054–2073
- Xiong J, Subramaniam S, Govindjee (1998) A knowledge-based three dimensional model of the Photosystem II reaction center of *Chlamydomonas reinhardtii*. *Photosynth Res* 56:229–254
- Young ID, Ibrahim M, Chatterjee R, Gul S, Fuller F, Koroidov S, Brewster AS, Tran R, Alonso-Mori R, Kroll T, Michels-Clark T, Laksmono H, Sierra RG, Stan CA, Hussein R, Zhang M, Douthit L, Kubin M, de Lichtenberg C, Vo Pham L, Nilsson H, Cheah MH, Shevela D, Saracini C, Bean MA et al. (2016) Structure of photosystem II and substrate binding at room temperature. *Nature* 540:453–457
- Zhang C, Chen C, Dong H, Shen J-R, Dau H, Zhao J (2015) A synthetic Mn_4Ca -cluster mimicking the oxygen-evolving center of photosynthesis. *Science* 348:690–693